

The Potential Effects of Garden Cress Seeds (*Lepidium Sativum L.*) on the Bone of Female Rats Suffering from Osteoporosis

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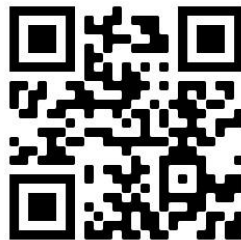
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Abstract

Osteoporosis is associated with a high rate of morbidity and mortality worldwide. This study was designed to investigate the probable effect of Garden Cress seeds (GCs) on the bone of osteoporotic female rats. Thirty healthy adult female albino rats (weigh, 200±10g) were used in this study. The rats were haphazardly divided into two main groups. The first main group (6 rats) was fed on a basal diet as a control negative group. The second main group (24 rats) fed on a basal diet containing 100mg prednisone acetate as a source of glucocorticoid/kg diet for two weeks to induce osteoporosis. One group was selected as a positive control group, three groups were fed on prednisone acetate diets containing GCs powder at the level of 2.5, 5, and 7.5%. The proximate chemical composition of GCs powder contained 2.75, 23.67, 22.81, 5.16, 8.42, and 37.19 g/100g for moisture, protein, fat, ash, crude fiber, and total carbohydrates respectively, while caloric value recorded 448.73 Kcal/100g. The minerals content were 309.21, 612.54, 341.10, and 5.74 mg/100g for calcium, phosphorus, magnesium, and zinc. The biological results showed a significant improvement ($P \leq 0.05$) in values of Ca and P in serum and femur bone of osteoporotic rats fed on a basal diet containing different ratios of GCs. Bone mineral content and bone mineral density also increased as compared to the control positive group. Kidney and

liver functions significantly ($P \leq 0.05$) improved compared to the positive control group. The histological examination of bone confirmed a gradual improvement in all treated groups. Peanut sweets were produced with proportions 2.5, 5, and 7.5% of GCs, sensory evaluation indicated that all peanut sweets samples have an acceptance greater than 75%. In conclusion, the current study indicates that GCs have improved osteoporosis biomarkers in rats and can be used up to 7.5% as a supplement in the diet of osteoporosis patients.

Keywords: osteoporosis, peanut sweets, calcium, phosphorus, bone mineral density.

Introduction

Osteoporosis is a chronic metabolic bone disease characterized by low bone mineral density (BMD), degradation of the bone structural, and increased fracture risk (Wang et al., 2017). Women over 45 years of age are the most vulnerable to femoral fractures, which are rising and pose a health and economic burden on society (Wu et al., 2018). Recent treatments of osteoporosis concentrate on slowing bone resorption and increasing bone remodeling (Wu et al., 2017). Pharmacological treatments that including calcium and vitamin D supplementation have been associated with a variety of side effects, especially with long-term usage (Hough et al., 2014).

Natural plant products as medication substitutes remain an important element of traditional medical regimens in developing nations (Veeresham, 2012). Garden cress (*Lepidium sativum L.*) is a fast-growing annual herb from the family *Cruciferae*, native to Egypt and west of Asia and is presently cultivated all over the world (Singh et al., 2015). Garden cress seeds (GCs) are an edible plant known as “Hab Al-Rashad” in Egypt, it can be used to fortify a variety of beverages and foods due to its excellent nutritional and functional qualities (Sakran et al., 2014).

GCs include 27% protein, fat ranging from 14 to 26%, carbohydrates from 35 to 54%, and 8% crude fiber. GCs carbohydrates are composed of 10% starch and 90% non-starch polysaccharides, GCs contain 20–25% oil, with α -linolenic acid accounting for 32–35% of

the totality, while the total essential amino acid content is 47.08% (**Chatoui et al., 2020**). They also include natural antioxidants that aid in the protection from oxidation and rancidity in the oil (**Zia-Ul- Haq et al., 2012**). GC seeds have a remarkable amount of iron, calcium, phosphorus, folic acid, vitamins C and A (**Lahiri and Rani, 2020**). Furthermore, the phytochemicals ingredients in seeds contain alkaloids, saponins, and flavonoids (**Manohar et al, 2012**).

GCs are used in several medicinal applications as a tonic for the immune system, in treating anemia (**Doke and Guha, 2014**), antidiarrheal (**Manohar et al., 2009**), antidiabetic (**Rahimi 2015**), antihypertensive, diuretic (**Maghrani et al., 2005**), antioxidant (**Yadav 2010**), anti-carcinogenic (**Al-Sheddi et al., 2016**), anti-hypercholesterolemia (**Al Hamedan 2010**), antibacterial, antifungal, anti-inflammatory (**Alqahtani et al., 2019**), anti-osteoporosis activity, normal contraction of the muscles for healthy of limbs and heart activity (**Jabeen et al., 2017**), and accelerate fracture healing (**Abdallah et al., 2020 and Shah et al., 2021**).

Moreover, GCs have a potential therapeutic effect against liver toxicity and hepato-carcinoma (**Ahmad et al., 2020**), also used in lactating mother's diets to enhance milk secretion during lactation, as well as to help regulate menstruation and thyroid hormone concentration (**Obeid, 2020 and Sciarrillo et al., 2018**). Additionally, **Mabrouk et al. (2020)** discovered that GCs oil improved the osteoporosis bone marker in both therapeutic and preventative methods. According to **Datta et al. (2011)**, GCs did not cause toxic effects even at 10% dietary level, as shown by the absence of any negative effects on growth or analytical parameters which means GCs are safe to consume in nutritional compositions.

Many people are still ignorant of the benefit of consuming GCs in their diets. Therefore, the present study aims to investigate the potential effect of GCs on the bone of osteoporotic rats, in addition to evaluating the sensory characteristics of peanut sweets supplemented with Garden cress seeds.

Materials and Methods

Materials

- Garden cress seeds (GCs) were purchased from Haraz Company for Agricultural Seeds, Perfumery and Medicinal Plants, Egypt.
- Casein, vitamins, minerals, cellulose, and choline chloride were purchased from El-Gomhoriya Company for Trading Drugs, Chemicals and Medical instruments, Cairo, Egypt.
- Prednisone acetate was purchase from a pharmacy in Damietta Governorate, Egypt.
- All ingredients used in peanut sweets formulation (Peanut seeds, sugar, corn syrup, vanilla flavor, and lemon) were obtained from the local market from Damietta Governorate, Egypt.

Methods

Preparation of Garden cress seeds

GCs were cleaned manually to remove adhering dirt, dust, and foreign particles along with shrunken and broken seeds. The seeds were roasted in a preheated iron skillet at a temperature of 128°C for 3 minutes leading to loss of raw flavor and production of a prominent aroma (**Vaishnavi and Gupta, 2020**). Roasted seeds were taken out from the skillet and spread on a tray for rapid cooling. After cooling, GCS were ground in electric milling. Packaging of roasted seeds flour was in air-tight plastic bags and stored in an air-sealed container till use.

Determination of gross chemical composition

The proximate major chemical constituents; moisture, ash, total protein, crude fiber, and fat in GCs powder were determined according to **A.O.A.C. (2000)**, while total carbohydrates were calculated by differences. Caloric value was calculated according the following equation: Caloric value = 4 (protein%+ carbohydrates%) + 9 (fat%).

Minerals content in GCs; calcium, phosphorus, magnesium, and zinc were measured using Perkin Elmer Atomic Absorption Spectrophotometer (Model 2380, Japan) according to the method of **A.O.A.C. (2010)**

Biological experiments design

The biological studies were carried out in accordance with the National Research Council's Institute of Laboratory Animal Resources, Commission on Life Sciences regulations (NRC, 2011). Thirty healthy adult female albino rats Sprague Dawley Strain weighing 200 ± 10 g per each were purchased from the Laboratory Animal Unit in the Nile Center of Experimental Research, Mansoura, Egypt. The rats were individually housed in wire cages under the normal laboratory condition and were fed on a basal diet for one week as an adaptation period. The basic diet was prepared according to the following formula as mentioned by (AIN, 1993), while the used vitamins and salt mixtures components were formulated according to Campbell, (1963) and Hegsted *et al.* (1941), respectively. After that, rats were randomly divided into two main groups, the first main group (6 rats) was fed on a basal diet, as a control negative group (C-). The second main group (24 rats) fed on a basal diet containing 100mg (prednisone acetate) as a source of glucocorticoid / kg diet for two weeks to induce osteoporosis according to Liao *et al.* (2003) The second main group was divided into four subgroups of 5 rats each as follows; subgroup 1: positive control (C+) fed on a prednisone acetate diet, subgroup 2: fed on a prednisone acetate diet containing GCs 2.5%, subgroup 3: fed on a prednisone acetate diet containing GCs 5%, and subgroup 4: fed on a prednisone acetate diet containing GCs 7.5%. Rats were maintained under standard conditions ($23\pm 2^{\circ}\text{C}$ temperature, $55\pm 5\%$ relative humidity, 12h light/12h dark cycle). The animals were fed diet and water ad-libitum for a period of 4 weeks. The diets consumed as well as body weights were recorded twice weekly.

At the end of the experiments, all rats were fasted up to 12 hours and then sacrificed. Blood samples were collected from the aorta. The blood samples were centrifuged, separated, and stored frozen at -20°C until further analysis.

Biochemical evaluation

Calcium and Phosphorus: Serum calcium and phosphorus samples were analyzed according to Baginski (1973) and Yee (1968), respectively. Calcium and phosphorus in the femur bone of the rats

were determined according to the method described by **Muynck and Vanhaecke, (2009)**.

Kidney functions: Serum levels of uric acid, urea nitrogen and creatinine were determined according to **Fossati *et al.* (1980)**, **Patton and Crouch (1977)**, and **Bohmer (1971)**, respectively.

Liver functions: Serum levels of aspartate aminotransferase (AST) and alanine aminotransferase (ALT) activities were measured according to **Tietz (1976)**, while Alkaline phosphates (ALP) activity was determined according to **Vassault *et al.* (1999)**.

Bone Mineral Density (BMD) and Bone Mineral Concentration (BMC) were both measured by Dual Energy X-ray Absorptiometry (DEXA) in the osteoporosis unit at the Egyptian Center for Medical and Scientific Research of Excellence.

Histological examination

Each rat's bones were removed, cleaned, weighed and specimens of right cortical bone were fixed with 10% neutral formalin solution till histopathologically examined according to **Bancroft and Cook, (1998)**.

Preparation of Peanut Sweets

Preparation of standard formula from peanut sweets (control) was produced by following the traditional method; Peanuts (100g), sugar (50g), corn syrup (25g), water (20ml), and lemon juice (5ml). Sugar and water were heated and made into syrup, then lemon juice was added while stirring the mixture till the temperature reached (150°C). The mixture was left boiling till the syrup turned glossy and thickened and the hard crack stage was reached. The corn syrup was added and the mixture stirred until completely homogeneous, then flavored with vanilla and kept away from the flame. Roasted and crushed peanuts were added to this syrup and were mixed thoroughly to cover the peanuts properly. The mixture was then transferred on a pre-greased surface and spread uniformly with the help of a roller, cut into square pieces, and put away to cool at room temperature. The other

experimental formulations were prepared by addition of powder from GC seeds with 2.5, 5, and 7.5%.

Sensory analysis

Peanut sweets samples were served in random order to ten panelists of staff members from Damietta University, Damietta, Egypt. Sensory attributes for color, taste, flavor, texture, appearance, and acceptance were evaluated using a 9- point hedonic scale (1= dislike extremely to 9 = like extremely) (IFT, 1981). Water was served for cleaning the mouth between samples.

Statistical Analysis:

Data obtained were statistically analyzed by SPSS computer software SPSS 2000. The results were expressed as mean \pm standard deviation (SD) and tested for significance using one-way analysis of variance ANOVA test, according to **Armitage and Berry, (1987)**.

Results and Discussion

Proximate chemical composition of Garden cress seeds

The proximate chemical composition of Garden cress seeds powder (GCs) was investigated on a dry weight basis and presented in table (1). It could be observed that GCs contained 2.75 ± 0.09 , 23.67 ± 0.05 , 22.81 ± 0.06 , 5.16 ± 0.13 , 8.42 ± 0.07 , and 37.19 g/100g for moisture, crude protein, fat, ash, crude fiber, and total carbohydrate, respectively. These findings revealed that seeds had a high amount of protein and low moisture content that is an indicator of stable quality and increased shelf life of seeds. Additionally, GCs had a high food energy content that reached 448.73 Kcal/100g for caloric value. On the other hand, GCs contained several minerals; 309.21 ± 0.14 , 612.54 ± 0.12 , 341.10 ± 0.06 , and 5.74 ± 0.03 mg for calcium, phosphorus, magnesium and zinc, respectively. The results showed that GCs are a good source of minerals that can protect bones.

**Table (1): Proximate chemical composition of GCs powder
(%On dry weight basis)**

Components	Ingredients
Moisture (g)	2.75 ±0.09
Protein (g)	23.67±0.05
Fat (g)	22.81±0.06
Ash (g)	5.16±0.13
Crude fiber (g)	8.42±0.07
Total Carbohydrate (g)	37.19
Caloric Value (Kcal)	448.73
Calcium (mg)	309.21±0.14
Phosphorus (mg)	612.54±0.12
Magnesium (mg)	341.10±0.06
Zinc (mg)	5.74±0.03

Each value represents the mean ± SD.

Feed intake, body weight gain% and femur bone weight relative to body weight %

Results of feed intake (FI), body weight gain% (BWG%), and femur bone weight/body weight% are summarized in table (2). It could be observed that FI (g/day/rat), BWG%, and femur bone weight/body weight% of the osteoporotic group (C+) decreased significantly ($p \leq 0.05$) when compared to the control negative group. That was due to the rats receiving a basal diet containing prednisone acetate, which reduced both appetite and body weight. Osteoporotic groups which fed on a prednisone acetate diet containing 2.5%, 5%, and 7.5% GCs showed a high gradual increase in FI, BWG%, and femur bone weight/body weight% as compared to the control (+) group. Statistical analysis showed significant positive correlations between treatments for FI, BWG%, and femur bone weight/body weight% by GCs. These results are in line with those reported by **Mali *et al.* (2007)** who found that *Lepidium Sativum* contains growth promoter factors as triterpenes, alkaloid, tannin, and coumarins. **Al Hamedan (2010)** indicated that rat groups with oral administration of garden cress seed extract showed a significantly lower value of weight gain.

Table (2): Effect of GCs on feed intake, Body weight gain% and femur bone weight/body weight% of female osteoporotic rats

Parameters Groups	Feed Intake g/day/rat	BWG%	Femur Weight/ body weight%
Control -	17.34 ^a ±1.56	20.70 ^a ±1.78	1.63 ^a ±0.07
Control +	11.21 ^c ±0.85	7.53 ^d ±1.22	0.84 ^d ±0.09
2.5% GCs	13.54 ^b ±1.15	9.14 ^c ±1.83	1.10 ^c ±0.06
5% GCs	14.32 ^b ±0.53	12.31 ^b ±1.69	1.28 ^b ±0.05
7.5% GCs	14.71 ^b ±1.21	13.83 ^b ±1.70	1.32 ^b ±0.06

GCs: Garden cress seeds

Means in the same column with different superscript letters are significantly different at $p \leq 0.05$.

Serum calcium and phosphorus, Femur bone calcium and phosphorus, bone mineral density, and bone mineral concentration

Calcium is the major component of the bone skeleton and possesses a significant role in osteoporosis prevention because its level is directly related to bone mineral density (BMD) and bone health (Hejaz et al., 2020). The calcium-to-phosphorus ratio, rather than the calcium or phosphorus level alone, is a deciding factor in bone validity and an osteoporosis prediction (Shakoor et al., 2014).

Results of feeding osteoporotic female rats on diets containing different levels of GC seeds on serum calcium and phosphorus, femur bone calcium and phosphorus, bone mineral density (BMD), and bone mineral concentration (BMC) were illustrated in the table (3). It could be noticed that untreated osteoporotic group (C+) recorded significant decreases in serum calcium and phosphorus (mmol/l) as compared them with the healthy control group (C-) (2.19 ± 0.21 and 1.42 ± 0.15 vs. 5.04 ± 0.31 and 3.12 ± 0.37 mmol/l, respectively), Treating osteoporotic groups with a prednisone acetate diet supplemented with 2.5, 5, and 7.5% GCs improved the levels of serum calcium and phosphorus in rats suffering from osteoporosis. The highest increase in serum calcium and phosphorus levels appeared in the osteoporotic group fed on a prednisone acetate diet supplemented with 7.5% of GCs which

recorded non-significant changes as compared to the control negative group.

Femur bone calcium and phosphorus of healthy rats (C-) fed on a basal diet increased than that of osteoporotic rats (C+) (50.2 ± 2.92 and 94.64 ± 2.50 vs. 30.62 ± 3.03 and 61.36 ± 4.66 mg/kg, respectively). Treating osteoporotic groups with a prednisone acetate diet supplemented with 5% and 7.5% GCs elevated the levels of calcium and phosphorus in femur bone of rats suffering from osteoporosis. The best result was recorded for the group of female rats which received a prednisone acetate diet containing 7.5% of GC seeds. These findings observed that GCs improved biochemical indicators of bones.

Bone loss is one of the most common adverse effects of glucocorticoid usage, even at low dosages. The major impact of glucocorticoids on bone is to suppress osteoblast activity, which results in a reduction in bone formation (De Nijs, 2008). Regarding the mean values of both bone mineral density (BMD) and bone mineral concentration (BMC), it could be noticed that employing prednisone acetate to induce osteoporosis led to a significant decline in BMD and BMC (g/cm^2) in the positive control group, as compared to the negative control group (0.09 ± 0.02 and 0.16 ± 0.03 vs. 0.17 ± 0.01 and 0.31 ± 0.02 g/cm^2 , respectively). The obtained results indicated that BMD and BMC increased gradually with the increasing levels of GC seeds. The best results of BMD and BMC of all tested groups were recorded for the group treated with 7.5% followed by 5% GC seeds. The results showed the positive effect of GC seeds in improving BMD and BMC due to their minerals content which stimulates the formation of bone osteoblasts and prevents bone resorption. Also, the Histopathological examination confirmed these results.

These results are in line with those reported by Bukhari *et al.* (2018) who indicated that *L. sativum* improves bone structure and reduces symptoms that occur with arthritis and body systems inflammation; where it was observed significant changes in BMD and improved T-Score at baseline after treatment with *L. sativum*. Al-Tamimi *et al.* (2020) mentioned that *Lepidium sativum* seeds elevated

the serum calcium level. According to **Mabrouk *et al.* (2020)** GC seeds oil increased mineral and vitamin D concentrations, and improved histological structure and bone thickness in rats suffering from osteoporosis. **Alharbi *et al.* (2021)** demonstrated that *L. sativum* seeds improve bone healing and could be used as a supplemental or alternative therapeutic osteogenic agent in the therapy of bone fractures.

Table (3): Effect of GCs on serum calcium and phosphorus, femur bone calcium and phosphorus, BMD and BMC of female osteoporotic rats

Parameters Groups	Serum Ca	Serum P	Femur bone Ca	Femur bone P	BMD	BMC
	mmol/L		mg/kg		g/cm ²	
Control -	5.04 ^a ±0.31	3.12 ^a ±0.37	50.2 ^a ±2.92	94.64 ^a ±2.50	0.17 ^a ±0.01	0.31 ^a ±0.02
Control +	2.19 ^d ±0.21	1.42 ^d ±0.15	30.62 ^c ±3.03	61.36 ^d ±4.66	0.09 ^d ±0.02	0.16 ^d ±0.03
2.5% GCs	3.13 ^c ±0.51	2.00 ^c ±0.50	31.15 ^c ±2.55	63.45 ^d ±5.32	0.13 ^c ±0.02	0.20 ^c ±0.02
5% GCs	4.12 ^b ±0.28	2.29 ^{bc} ±0.37	40.72 ^b ±2.19	72.33 ^c ±7.28	0.14 ^{bc} ±0.01	0.25 ^b ±0.01
7.5% GCs	4.64 ^{ab} ±0.41	2.86 ^{ab} ±0.61	46.65 ^a ±3.27	83.01 ^b ±7.11	0.16 ^{ab} ±0.01	0.27 ^b ±0.02

GCs: Garden cress seeds, BMD: Bone Mineral Density, BMC: Bone Mineral Concentration.
Means in the same column with different superscript letters are significantly different at $p \leq 0.05$

Kidney function

According to **Jeon (2008)**, the kidney regulates calcium excretion by reabsorbing approximately 95% of filtered calcium through the renal tubules. Data presented in Table (4) showed the effect of GC seeds on serum uric acid, urea nitrogen, and creatinine of female rats suffering from osteoporosis. The mean values \pm SD of uric acid, urea nitrogen, and creatinine (mg/dl) of the osteoporotic group (C+) were significantly ($P \leq 0.05$) increased as compared with the intact group (2.51 ± 0.15 , 60.31 ± 4.54 and 1.55 ± 0.09 vs. 1.57 ± 0.07 , 30.42 ± 1.55 and 0.52 ± 0.04 mg/dl, respectively). Treating osteoporotic groups with a prednisone acetate diet supplemented with 5% and 7.5% GCs decreased the levels of serum uric acid, urea nitrogen, and creatinine of

rats suffering from osteoporosis, while the osteoporotic group of rats which treated with the low level of GCs didn't differ significantly in serum urea nitrogen and creatinine as compared with the positive control group. The results showed that the biologically active phytochemicals ingredients in garden cress could improve kidney functions in osteoporotic rats.

The present study is in harmony with **Al Hamedan (2010)** who found that oral administration of garden cress seeds extract and powder showed significantly lower values for serum urea and creatinine in rats. Similarly, **Nilesh et al. (2010)** discovered that administering *L.sativum* extract reduced serum creatinine and urea levels substantially. **Sakran et al. (2014)** reported that glucosinolates and other compounds in *L. sativum* seed are responsible for protecting cells of the liver and kidney against various toxins. Also, **Halaby et al. (2015)** mentioned that the diets fortified at 5% and 10% seeds powder of *L. sativum* helped to improve kidney and liver function compared with positive control groups.

Table (4): Effect of GCs on serum kidney functions of female osteoporosis rats

Parameters Groups	Uric acid	Urea nitrogen	Creatinine
	mg/dl		
Control -	1.57±0.07 ^c	30.4±2±1.55 ^d	0.52±0.04 ^d
Control +	2.51±0.15 ^a	60.31±4.54 ^a	1.55±0.09 ^a
2.5% GCs	2.32±0.20 ^b	56.98±4.66 ^a	1.49±0.05 ^a
5% GCs	1.99±0.12 ^c	49.82±3.09 ^b	1.17±0.04 ^b
7.5% GCs	1.79±0.08 ^d	43.68±3.23 ^c	0.89±0.05 ^c

GCs: Garden cress seeds

Means in the same column with different superscript letters are significantly different at $p \leq 0.05$.

Liver enzymes activity

Results illustrated in Table (5) revealed the effect of GC seeds on serum liver functions of osteoporotic female rats. The mean values \pm SD of serum AST, ALT, and ALP (u/L) significantly increased

($P \leq 0.05$) in the positive control group as compared to the negative control group (133.47 ± 3.14 , 57.23 ± 3.71 and 177.84 ± 4.52 vs. 70.18 ± 4.41 , 26.61 ± 2.77 and 81.37 ± 2.21 u/L, respectively), because of nourishing prednisone acetate diet. Feeding osteoporotic groups on a prednisone acetate diet containing different levels of GCs showed a significant decrease in the mean values of serum AST, ALT and ALP, as compared to the positive control group. The best result of liver enzymes activity was recorded for the group treated with 7.5% GC seeds.

These results were in line with those found by **Al-Asmari et al. (2015)** who showed that *L. sativum* seeds ethanolic extract improves liver enzymes significantly. Similarly, **Raish et al. (2016)** found that supplementation with *L. sativum* seeds extract ameliorated liver enzymes activity and reduced hepatic injuries and structural damage by decreasing oxidative stress, inflammation, and apoptosis in the liver had.

Table (5): Effect of GCs on serum liver enzymes activity of female osteoporosis rats

Parameters Groups	AST	ALT	ALP
	u/L		
Control -	70.18 ± 4.41^c	26.61 ± 2.77^d	81.37 ± 2.21^c
Control +	133.47 ± 3.14^a	57.23 ± 3.71^a	177.84 ± 4.52^a
2.5% GCs	119.53 ± 4.34^b	53.09 ± 6.37^a	163.88 ± 3.76^b
5% GCs	107.77 ± 4.57^c	46.31 ± 2.08^b	153.59 ± 3.07^c
7.5% GCs	98.08 ± 4.42^d	38.31 ± 4.19^c	146.73 ± 3.59^d

GCs: Garden cress seeds

Means in the same column with different superscript letters are significantly different at $p \leq 0.05$.

Histological Examination

Fig. (1) showed the histopathological examination of the femur right bone. Microscopically, the bone of rats from C- group which fed on a basal diet revealed the normal histological structure of trabecular


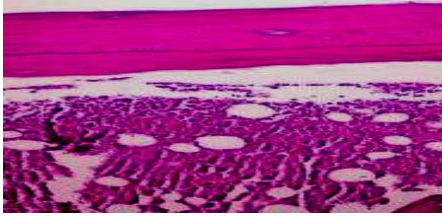
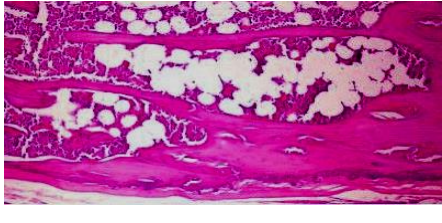
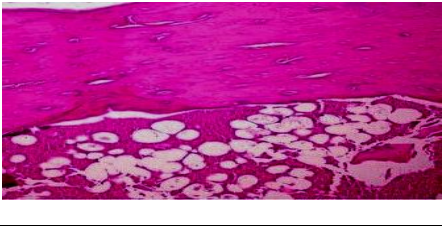
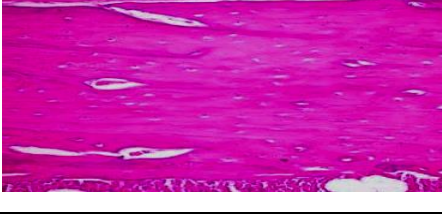
	<p>Photo (1): Control - Normal histological structure of trabecular bone (H & E x 200).</p>
	<p>Photo (2): Control+ Reduction in bone mass and the medullary cavity becomes enlarged (H & E x 200)</p>
	<p>Photo (3): 2.5% GCS Focal necrosis in bone cortex, few bone trabecular, and enlarged medullary cavity. (H & E x 200)</p>
	<p>Photo (4): 5% GCS Thick cortical bone and massive proliferation of osteoblasts, while some cavity still present. (H & E x 200)</p>
	<p>Photo (5): 7.5% GCS Obvious normal bone structure. (H & E x 200).</p>

Fig (1): Histological examination of femur bone structure

bone (Photo1). While, the bone of rats suffering from osteoporotic C+ group showed a reduction in bone mass and the medullary cavity became enlarged (photo 2). In this respect, (Photos 3, 4 & 5) showed a gradual improvement of bone tissue, thick cortical bone and massive proliferation of osteoblasts in osteoporotic rats that received a prednisone acetate diet containing different ratios of GCs. The results

agreed with **Mabrouk *et al.* (2020)** who found that consumption of GCs oil leads to enhancing the histological structure and bone thickness of osteoporotic rats.

Sensory properties of prepared peanut sweets with GCs

Peanut sweets are a popular product among all population groups in Egypt, especially when celebrating the Prophet's birthday. This kind of sweets can be made out of different nuts, the current study had chosen peanuts to make peanut sweets.

Sensory evaluation is a crucial indicator of potential consumer preferences. Table (6) presented the sensory characteristics of prepared peanut sweets with garden cress seeds. It could be observed that all formulations were acceptable in all sensory evaluation attributes (color, taste, flavor, texture, and general acceptability), there were no significant differences ($P \leq 0.05$) between the samples supplemented with 2.5% and 5% of GCs when compared with the control sample. While the flavor of the sample which contained 5% GCs slightly decreased as compared to the control peanut sweets sample. The addition of high levels of GCs by 7.5% caused significant changes ($P \leq 0.05$) in the same sensory evaluation attributes as compared to the control sample, but the sample was still acceptable. General acceptance score ranged from 8.24 to 6.78 by about 91.55 to 76.33%. It means that garden cress seeds can be used in the preparation of peanut sweets and any other product up to 7.5%.

These results are in harmony with **Umesha *et al.* (2015)** who indicated that biscuits prepared by supplementing garden cress oil at 5g/100 g by replacing fat were acceptable. **Alshehry (2019)** found that biscuits fortified with garden cress powder till 7.5 percent acceptance are the most popular among panelists.

Table (6): Sensory characteristics of prepared Sweet Peanuts with GCs

Sensory Characteristics	Control 0%	Sweet peanuts with GCs		
		2.5%	5%	7.5%
Color	8.53 ^a ±.262	8.47 ^a ±.371	8.41 ^a ±.200	8.08 ^b ±.101
Taste	8.64 ^a ±.281	8.57 ^{ab} ±.301	8.31 ^{ab} ±.383	7.93 ^b ±.380
Flavor	8.81 ^a ±.360	8.35 ^{ab} ±.281	7.68 ^{bc} ±.420	7.44 ^c ±.350
Texture	8.38 ^a ±.331	8.19 ^a ±.311	7.86 ^{ab} ±.321	7.12 ^b ±.510
General Acceptance	8.24 ^a ±.210	8.12 ^a ±.248	7.67 ^{ab} ±.385	6.87 ^c ±.231

GCs: Garden cress seeds

Means with different letter within the same row are significantly different at $P \leq 0.05$

Conclusion

According to the findings, Garden Cress seeds are a good source of minerals and a variety of efficient prophylactic chemicals which significantly improved the osteoporosis bone markers. Garden Cress seeds may be utilized in many products as a functional food to reduce the risk of osteoporosis. Eventually, Garden Cress seeds can be applied up to 7.5% in functional foods.

References

- A.O.A.C. (2000).** Official Methods of Analysis (17th ed.) Arlington, VA: Association of Official Analytical Chemists, AOAC International.
- A.O.A.C. (2010).** Official Methods of Analysis Association of Official Analytical Chemists International, 19th ed., Washington, D. C., USA.
- Abdallah, H.M.; Farag, M.A.; Algandaby, M.M.; Nasrullah, M.Z.; Abdel-Naim, A.B.; Eid, B.G. and Malebari, A.M. (2020).** Osteoprotective activity and metabolite fingerprint via UPLC/MS and GC/MS of *Lepidium sativum* in ovariectomized Rats. *Nutrients*, 12(7): 2075.
- Ahmad, A.; Nabi, R.; Mishra, A. and Ahmad, I. Z. (2020).** A Panoramic Review on *Lepidium sativum* L. Bioactives as Prospective Therapeutics. *Drug Research*, 71(05):233-242.
- AIN (1993).** American institute of nutrition purified diet for laboratory rodent: final report to *Journal Nutrition*, 123: 1939-1951.
- Al Hamedan, W. (2010).** Protective effect of *Lepidium sativum* L. seeds powder and extract on hypercholesterolemic rats. *Journal of American Science*, 6(11): 873-879.
- Al-Asmari, A.K.; Athar, M.T.; Al-Shahrani, H.M.; Al-Dakheel, S.I. and Al-Ghamdi, M.A. (2015).** Efficacy of *Lepidium sativum* against carbon tetra chloride induced hepatotoxicity and determination of its bioactive compounds by GCMS. *Toxicology reports*, 2:1319-1326.
- Alharbi, F.H.; Baothman, O.A.S.; Zamzami, M.A.; Abou Gabal, H.H.; Khoja, S.M.; Karrouf, G. and Abo-Golayel, M.K. (2021).** Garden cress (*Lepidium sativum* L.) seeds enhancing osteogenesis postinduced-bone fracture. *Pharmacognosy Magazine*, 17(73): 170-178.

- Alqahtani, F.Y.; Aleanizy, F.S.; Mahmoud, A.Z.; Farshori, N.N.; Alfaraj, R.; Al-Sheddi, E.S. and Alsarra, I.A. (2019).** Chemical composition and antimicrobial, antioxidant, and anti-inflammatory activities of *Lepidium sativum* seed oil. Saudi journal of biological sciences, 26(5):1089-1092.
- Al-Sheddi, E.S.; Farshori, N.N.; Al-Oqail, M.M.; Musarrat, J.; Al-Khedhairi, A.A. and Siddiqui, M.A. (2016).** Protective effect of *Lepidium sativum* seed extract against hydrogen peroxide-induced cytotoxicity and oxidative stress in human liver cells (HepG2). Pharmaceutical biology, 54(2):314-321.
- Alshehry, G.A. (2019).** Technological and sensory characteristics of biscuits fortified with garden cress (*Lepidium sativum*) seeds. Life Science Journal, 16(8):28-35.
- Al-Tamimi, A.H.S.; Radman, B.A.M.; Al-Dhuraibi, K.; Al-Naghi, N. and Qasim, M. (2020).** Effect of *Lepidium sativum* seeds on Serum and Urine Calcium in Male Albino Rats. Albaydha University Journal, 2(3):123-128.
- Armitage, P. and Berry, G. (1987).** Statistical Method in Medical Research. Blackwell. Oxford. UK. pp. 93-213.
- Baginski, E.S. (1973).** Method of calcium determination. Clin.chem. Acta, 46:49.
- Bancroft, J.D. and Cook, H.C. (1998).** Manual of histotechnological techniques. Edited by: Churchill Livingstone, New York, pp. 243-274.
- Bohmer, H. B.U.M. (1971).** Micro-determination of creatinine. Clin. Chem. Acta, 32:81-85.
- Bukhari, H.M.; Wahb, H.M.; Header, E.A.; Sharkar, A.A.; Alsaegh, N.A. and Bukhari, S.M. (2018).** Effects of *Lepidium Sativum* (Tuffa) on Bone Mineral Density and Symptoms in Female University Students in Makkah, Saudi Arabia: Single-Blind Randomized Trail. American Journal of Food Science and Nutrition Research, 5(3):48-54.

- Campbell, J.A. (1963).** Methodology of protein evaluation, PAG. *Nutrition. Document R*, 10, New York.
- Chatoui, K.; Harhar, H.; El Kamli, T. and Tabyaoui, M. (2020).** Chemical composition and antioxidant capacity of *Lepidium sativum* seeds from four regions of Morocco. *Evidence-Based Complementary and Alternative Medicine*, 4:1-7.
- Datta, P.K.; Diwakar, B.T.; Viswanatha, S.; Murthy, K.N. and Naidu, K. A. (2011).** Original Report Safety evaluation studies on Garden cress (*Lepidium sativum L.*) seeds in Wistar rats. *International Journal of Applied Research in Natural Products*, 4(1):37-43.
- De Nijs, R.N. (2008).** Glucocorticoid-induced osteoporosis: a review on pathophysiology and treatment options. *Minerva Med.*, 99(1):23-43.
- Doke, S. and Guha, M. (2014).** Garden cress (*Lepidium sativum L.*) Seed-An Important Medicinal Source: A Review. *Journal of Natural Product and Plant Resources*, 4(1)69-80.
- Fossati, P.; Prencipe, L. and Berti, G. (1980).** Enzymatic colorimetric method of determination of uric acid in serum. *Clin. Chem*, 26(2):227-273.
- Halaby, M.S.; Farag, M.H. and Mahmoud, S.A. (2015).** Protective and curative effect of garden cress seeds on acute renal failure in male albino rats. *Middle East Journal of Applied Sciences*, 5(2):573-586.
- Hegested, D.M.; Mills, R.C.; Elvehjen, C.A. and Hart, E.B. (1941).** Salt mixture. *Journal of Biological Chemistry*, 138:459.
- Hejazi, J.; Davoodi, A.; Khosravi, M.; Sedaghat, M.; Abedi, V.; Hosseinverdi, S. and Shojaie, L. (2020).** Nutrition and osteoporosis prevention and treatment. *Biomedical Research and Therapy*, 7(4): 3709-3720.

- Hough, F.S.; Brown, S.L.; Cassim, B.; Davey, M.R.; de Lange, W.; de Villiers, T.J. and Pettifor, J.M. (2014).** The safety of osteoporosis medication. South African Medical Journal, 104(4): 279-282.
- Institute of Food Technologists IFT (1981):** Sensory evaluation guide for testing food and beverage products. Journal of Food Science, 35(11):50-59.
- Jabeen, A.; Rani, S.; Ibrahim, M. and Mohammad, A.S. (2017).** A review on *Lepidium sativum*. Indo American Journal of Pharmaceutical Sciences, 4(8): 2223-2227.
- Jeon, U.S. (2008).** Kidney and calcium homeostasis. Electrolyte & Blood Pressure, 6(2): 68-76.
- Lahiri, B. and Rani, R. (2020).** Garden Cress Seeds: chemistry, medicinal properties, application in dairy and food industry: A Review. Emergent Life Sciences Research, 6(2):1-4.
- Liao, J.M.; Li, Q.N.; Wu, T.; Hu. B.; Huang, L.F.; Li, Z.H.; Zhao, W.D. Zhang M.C. and Zhong S.Z. (2003).** Effects of prednisone on bone mineral density and biomechanical characteristics of the femora and lumbar vertebrae in rats. Institute of Clinical Anatomy, First Military Medical University, Guangzhou 510515, China. Di 1 jun yi da xue xue bao= Academic Journal of the First Medical College of PLA, 23(2):97-100.
- Mabrouk, D.M.; Ibrahim, F.M.; Abdel-Aziem, S.H. and Sharaf, H.A. (2020).** Therapeutic and Prophylactic Efficacy of Garden Cress Seed Oil against Osteoporosis in Rats. Jordan Journal of Biological Sciences, 13(2):237-245.
- Maghrani, M.; Zeggwagh, N.A.; Michel, J.B. and Eddouks, M.(2005).** Antihypertensive effect of *Lepidium sativum* L. in spontaneously hypertensive rats. Journal of Ethnopharmacology, 100 (1-2):193-197.
- Mali, R.G.; Mahajan, S.G. and Mehta, A.A. (2007).** *Lepidium sativum* (Garden cress): a review of contemporary literature and

medicinal properties. *Oriental Pharmacy and Experimental Medicine*, 7(4): 331-335.

Manohar, D.; Shylaja, H.; Viswanatha, G.L. and Rajesh, S. (2009). Antidiarrheal activity of methanolic extracts of *Lepidium sativum* in rodent. *Journal of Natural Remedies*, 9(2):197–201.

Manohar, D.; Viswanatha, G.L.; Nagesh, S.; Jain, V. and Shivaprasad, H.N. (2012). Ethnopharmacology of *Lepidium sativum* Linn (Brassicaceae): a review. *International Journal of Phytotherapy Research*, 2(1): 1-7.

Muynck, D. and Vanhaecke, F. (2009). Development of a method based on inductively coupled plasma-dynamic reaction cell-mass spectrometry for the simultaneous determination of phosphorus, calcium and strontium in bone and dental tissue. *Spectrochimica Acta Part B: Atomic Spectroscopy* 64(5): 408-415.

Nilesh, S.; Amit, J.; Vaishali, U.; Sujit, K.; Sachin, K. and Ravindra, P. (2010). Protective effect of *Lepidium sativum* against doxorubicin-induced nephrotoxicity in rats. *Research Journal of Pharmaceutical Biological and Chemical Sciences*, 1(3):42-49.

NRC, National Research Council (2011). Guide for the care and use of laboratory animals. Guide for the care and use of laboratory animals. Washington, DC, USA: National Academies Press.

Obeid, A.K.; Al-Bazii, S. J. and Al-masoudi, F.J. (2020). Mammagensis effect of *lepidium sativum* seeds (garden cress) in mammary gland growth and development during three physiological stage in female rats. *EurAsian Journal of BioSciences*, 14(1):2273-2278.

Patton, C.J. and Crouch, S.R. (1977). Enzymatic colorimetric method to determine urea in serum. *Anal. Chem*, 49:464-469.

- Rahimi, M. (2015).** A review: antidiabetic medicinal plants used for diabetes mellitus. *Bull. Environ. Pharmacol. Life Sciencs*, 4(2):163-180.
- Raish, M.; Ahmad, A.; Alkharfy, K.M.; Ahamad, S.R.; Mohsin, K.; Al-Jenoobi, F.I. and Ansari, M.A. (2016).** Hepatoprotective activity of *Lepidium sativum* seeds against D-galactosamine/ lipopolysaccharide induced hepatotoxicity in animal model. *BMC complementary and alternative medicine*, 16(1):1-11.
- Reeves, B.G.; Nielson, F.H. and Fahmy, G.C. (1993).** Reports of the American institute of nutrition, Adhoc-Willing Committee on the reformulation of the AIN 93. Rodent diet. *Journal of Nutrition*, 123(11): 1939-1951.
- Sakran, M.; Selim, Y. and Zidan, N. (2014).** A new isoflavonoid from seeds of *Lepidium sativum* L. and its protective effect on hepatotoxicity induced by paracetamol in male rats. *Molecules*,19(10): 15440-15451.
- Sciarrillo, R.; Guarino, M. and Guarino, C. (2018).** Pharmacological activity of ethanolic extract *Lepidium sativum* linn. seeds on thyroid hormones in male rats. *International Journal of Pharmaceutical Sciences and Research*, 9(4): 1699-1704.
- Shah, M.B.; Dudhat, V.A. and Gadhvi, K.V. (2021).** *Lepidium sativum*: A potential functional food. *Journal of Ayurvedic and Herbal Medicine*,7(2):140-149.
- Shakoor, S.; Ilyas, F.; Abbas, N.; Mirza, M. A. and Arif, S. (2014).** Prevalence of osteoporosis in relation to serum calcium and phosphorus in aging women. *Journal Global Innovation in Agriculture and Social Science*, 2(2):70-75.
- Singh, C.S.; Paswan, V.K. and Naik, B. (2015).** Exploring potential of fortification by garden cress (*Lepidium sativum* L.) seeds for development of functional foods—A Review. *Indian Journal of*

Natural Products and Resources (IJNPR)[Formerly Natural Product Radiance (NPR)], 6(3):167-175.

Tietz, N.W. (1976). Fundamentals of clinical chemistry WB Saunders Co. Philadelphia PA, 47-51.

Umesha, S.S.; Manohar, R.S.; Indiramma, A.R.; Akshitha, S. and Naidu, K.A. (2015). Enrichment of biscuits with microencapsulated omega-3 fatty acid (Alpha-linolenic acid) rich Garden cress (*Lepidium sativum*) seed oil: Physical, sensory and storage quality characteristics of biscuits. LWT-Food Science and Technology, 62(1):654-661.

Vaishnavi, R.G. and Gupta, R. (2020). Effect of processing treatments on nutritional profile of garden cress (*Lepidium sativum* L.) seeds. International Journal Chemical Studies, 8(4):2831-2835.

Vassault, A.; Grafmeyer, D.; Graeve, J.; Cohen, R.; Beaudonnet, A.; and Bienvenu, J. (1999). Quality specifications and allowable standards for validation of methods used in clinical biochemistry. In Annales de biologie clinique, 57(6): 685-95.

Veeresham, C. (2012). Natural products derived from plants as a source of drugs. Journal of advanced pharmaceutical technology & research, 3(4):200-201.

Wang, Y.G.; Jiang, L.B. and Gou, B. (2017). Protective effect of vanillic acid on ovariectomy-induced osteoporosis in rats. African Journal of Traditional, Complementary and Alternative Medicines, 14(4): 31-38.

Wu, C.H.; Kao, I.J.; Hung, W.C.; Lin, S.C.; Liu, H.C.; Hsieh, M.H. and Yang, R.S. (2018). Economic impact and cost-effectiveness of fracture liaison services: a systematic review of the literature. Osteoporosis International, 29(6):1227-1242.

Wu, L.; Ling, Z.; Feng, X.; Mao, C. and Xu, Z. (2017). Herb medicines against osteoporosis: active compounds & relevant

biological mechanisms. Current topics in medicinal chemistry, 17(15):1670-1691.

Yadav, Y.C., Srivastav, D.N., Seth, A.K., Saini, V., Balaraman, R. and Ghelani, T.K. (2010). In vivo antioxidant potential of *Lepidium sativum* L. seeds in albino rats using cisplatin induced nephrotoxicity. International Journal of Phytomedicine, 2(3): 292-298.

Yee, H.Y. (1968). Dietary Calcium: adequacy a vegetarian diet. Am. J. Clin. Nutr., 59:12385.

Zia-Ul-Haq, M.; Ahmad, S.; Calani, L.; Mazzeo, T.; Del Rio, D.; Pellegrini, N. and De Feo, V. (2012). Compositional study and antioxidant potential of *Ipomoea hederacea* Jacq. and *Lepidium sativum* L. seeds. Molecules, 17(9):10306-10321.

التأثيرات المحتملة لبذور حب الرشاد على عظام إناث الفئران المصابة بهشاشة العظام

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الملخص

ترتبط هشاشة العظام بارتفاع معدل انتشار الامراض والوفيات من جميع أنحاء العالم. أجريت هذه الدراسة لمعرفة التأثيرات المحتملة لبذور حب الرشاد على عظام إناث الفئران المصابة بهشاشة العظام. استخدمت الدراسة ثلاثون أنثى فأر من نوع الألبينو (200±10جم). قسمت الفئران إلى مجموعتين رئيسيتين، المجموعة الرئيسية الاولى (6 فئران) أصحاء تم تغذيتهم على الوجبة الاساسية كمجموعة ضابطة سالبة. تمت تغذية المجموعة الرئيسية الثانية (24 فأر) على غذاء اساسي يحتوي على 100مجم بريدنيزون أسيتات كمصدر للجلوكوكورتيكويد / كجم لمدة أسبوعين للحث على احداث هشاشة العظام. تم اختيار مجموعة واحدة منهم كمجموعة ضابطة موجبة، اما المجموعات (3 و 4 و 5) تم تغذيتها على وجبة بريدنيزون أسيتات تحتوي على مسحوق من بذور حب الرشاد بنسبة 2.5 و 5 و 7.5%. أظهرت النتائج أن التركيب الكيميائي لمسحوق بذور حب الرشاد يحتوي على 2.75 و 23.67 و 22.81 و 5.16 و 8.42 و 37.19 جم / 100 جم لكل من الرطوبة والبروتين والدهون والرماد والألياف الخام وإجمالي الكربوهيدرات على التوالي، بينما سجلت قيمة السعرات الحرارية 448.73 كيلو كالوري / 100 جم. وبلغ محتوى المعادن 309.21، 612.54، 341.10، 5.74 ملجم / 100 جم من الكالسيوم والفوسفور والمغنيسيوم والزنك على التوالي. أظهرت نتائج الدراسة البيولوجية تحسناً معنوياً ($P \leq 0.05$) في قيم كل من الكالسيوم والفوسفور في مصل وعظم الفخذ في الفئران المصابة بهشاشة العظام والتي تغذت

على نظام غذائي أساسي يحتوي على نسب مختلفة من مسحوق بذور حب الرشاد، كما لوحظ زيادة كل من كثافة ومحتوى المعادن بالعظام مقارنة بالمجموعة الضابطة الموجبة. تحسنت وظائف الكلى والكبد معنوياً ($P \leq 0.05$) مقارنة بالمجموعة الضابطة الموجبة. أظهرت نتائج الفحص الهستولوجي للعظام تحسناً تدريجياً في أنسجتها. تم عمل حلوي الفول السوداني بالنسب 2.5 و 5 و 7.5% من مسحوق بذور حب الرشاد، وأشارت نتائج التقييم الحسي إلى أن جميع عينات حلوي الفول السوداني قد حظيت بقبول أكبر من 75%. خلصت نتائج الدراسة الحالية الي أن مسحوق بذور حب الرشاد قد حسن من مضاعفات الاصابة بهشاشة العظام في الفئران، كما يمكن استخدامه بنسبة تصل إلى 7.5% كمكمل في النظام الغذائي لمرضى هشاشة العظام.

الكلمات المفتاحية: هشاشة العظام- حلوي الفول السوداني- بذور حب الرشاد -الكالسيوم- الفسفور - كثافة المعادن بالعظام.